

Monte Carlo Simulations of Protein Folding using Lattice Models

Ryan Cheng^{1,2} and Kenneth Jordan^{1,3}

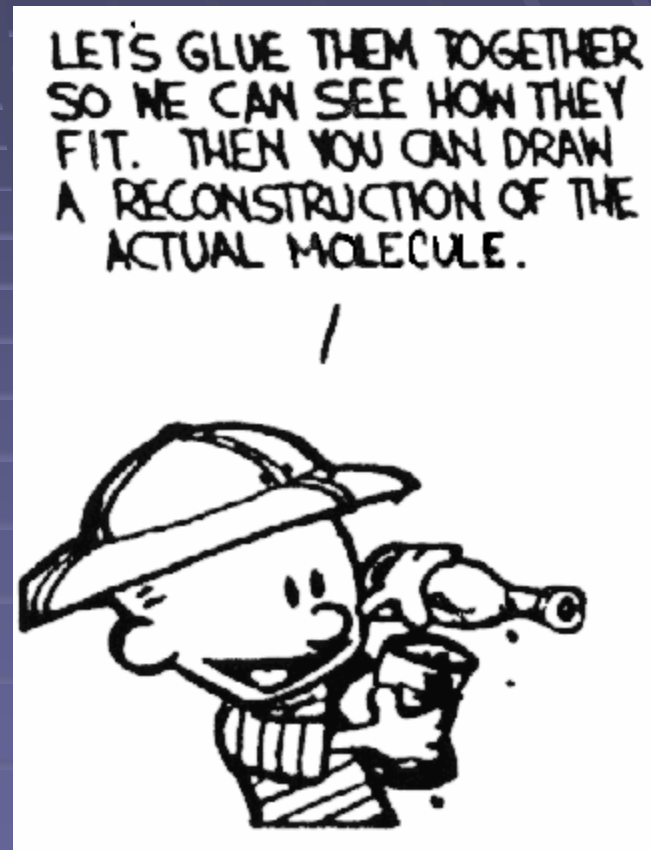
¹Bioengineering and Bioinformatics Summer Institute, Department of Computational Biology, University of Pittsburgh, Pittsburgh, PA 15261

²Department of Chemistry, Carnegie Mellon University, Pittsburgh, PA 15213

³Department of Chemistry, University of Pittsburgh, Pittsburgh PA, 15261

Goals

- Investigate folding of peptide sequences through simulation
- Gain insight on computational algorithms in protein folding
 - Random walk
 - MC-based Simulated Annealing

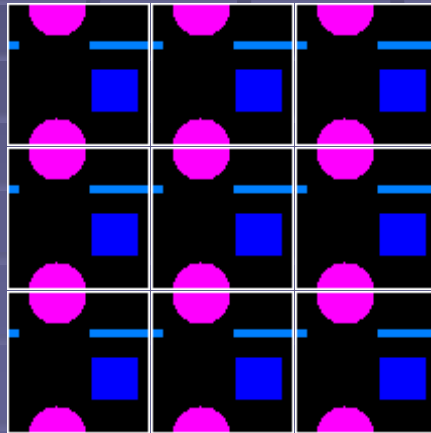


Introduction

- Protein native state
 - Direct relationship between conformation and biological function
 - Conformation located at global energy minimum on complex energy landscape
 - Methods of interest include computational techniques for optimization
 - “Levinthal’s Paradox”
 - Need for simplifying models (i.e., lattice models)

Lattice Model

- Two-dimensional square lattice
- Periodic boundary conditions—“Infinite lattice”



Lattice Model

- HP-lattice Model

- Hydrophobic effect assumed to be driving force in protein folding
- Amino acid monomers are modeled as being either hydrophobic (H) or polar (P)
 - Three possible interaction energies: E_{HH} , E_{HP} , and E_{PP}
- Energy function:

$$H = \sum_{i < j} E_{p_i p_j} \delta(r_i - r_j)$$

- $\delta(r_i - r_j) = 1$ if monomers r_i and r_j are adjacent non-bonded nearest neighbors and 0 otherwise

Simulated Annealing

- Based on idea of cooling molten material to form a perfect crystal
- Performed from effectively high temperature and cooled to frozen state
- Utilizes Metropolis Monte Carlo to minimize energy function
 - Moves accepted if:
 - 1) $\Delta E < 0$
 - 2) $\text{Random}[0, 1] \leq \text{Exp}[-\Delta E/T]$

Methodology

- Artificial peptide sequences:
 - SeqA used for simulation of alpha helices
(-Ala-Leu-Ser-Ser-Ala-Ala-Ser-)_n
(- H - H - P - P - H - H - P -)_n
20 Monomer Seq-A analyzed
 - SeqB used for simulation of beta sheets
(-Val-Ser-)_n
(- H - P -)_n
10 Monomer Seq-B analyzed

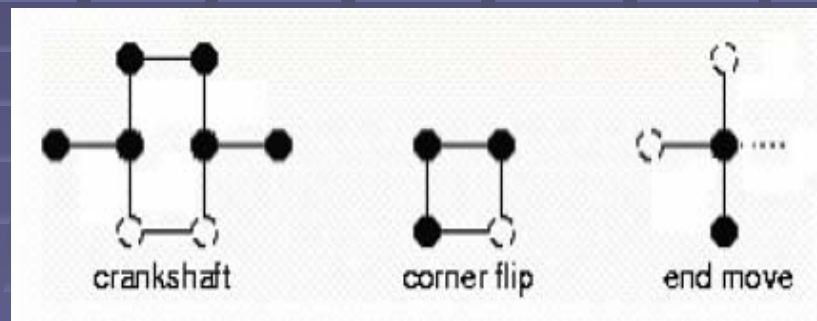
Methodology

- Self-avoiding random walk to generate peptide chain of length n
- Simulated annealing: 10,000 Metropolis iterations per temperature

	T_{start}	T_{stop}	T_{step}
1	100	50	-5
2	49	25	-1
3	24.5	10	-0.5
4	9.75	7	-0.25
5	6.9	0.05	-0.05

Methodology

- Metropolis reconfiguration based on Verdier-Stockmayer algorithm
 - End rotation, Kink jump, and crankshaft



- HP Interaction energies:
 - $E_{HH} = -3$ (most favorable)
 - $E_{HP} = -1.2$
 - $E_{PP} = 0$

Analysis

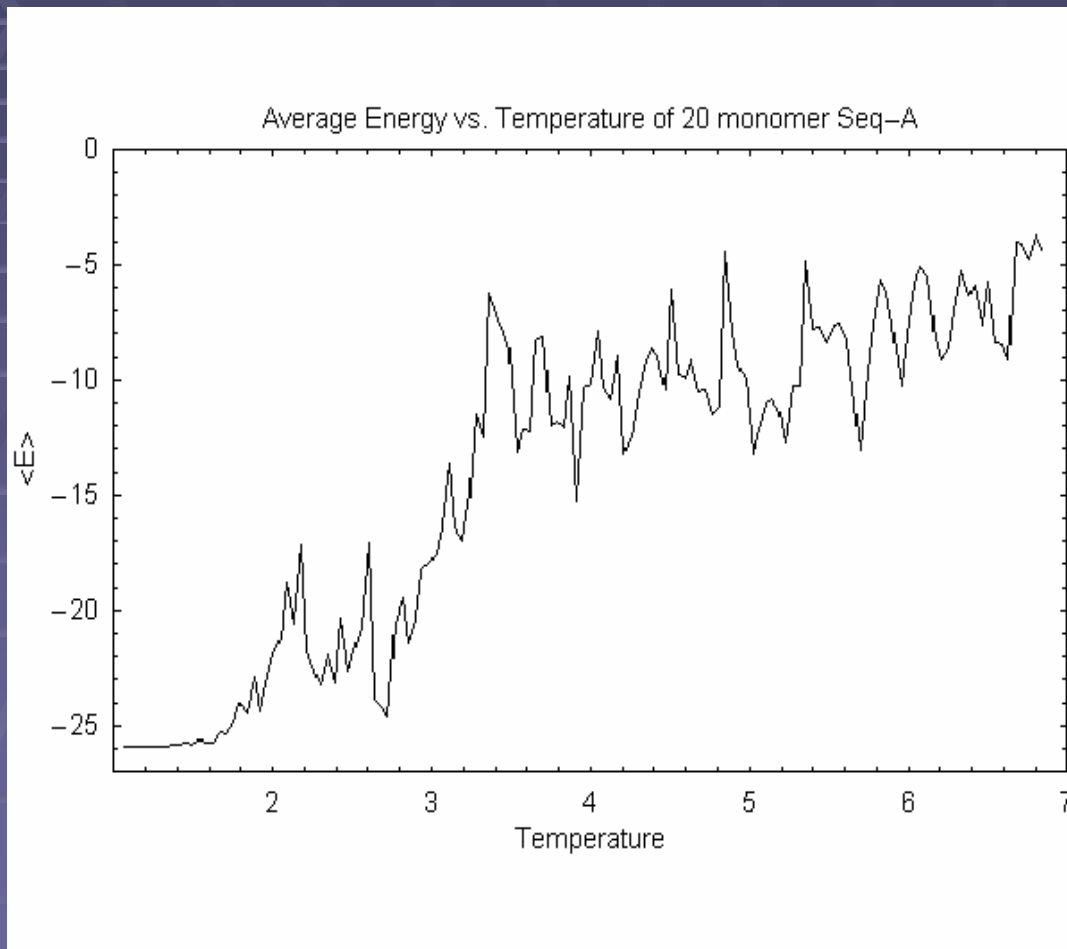
- $\langle E \rangle_T$ and $\langle E^2 \rangle_T$ calculated at each temperature
- Heat capacity calculated:

$$C(T) \propto \frac{\langle E^2 \rangle_T - \langle E \rangle_T^2}{T^2}$$

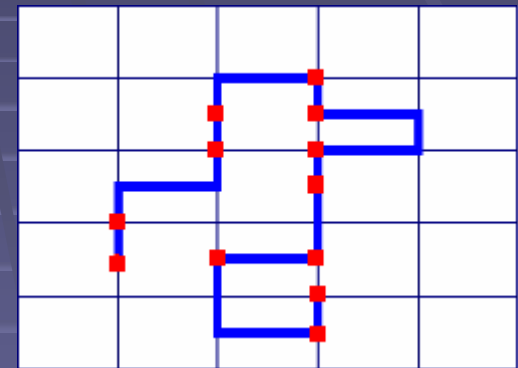
- Melting transition temperature observed
- Low energy conformation obtained

Results: 20 Monomer Seq-A

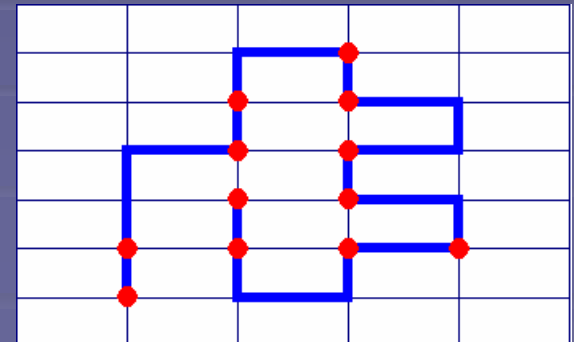
(H-H-P-P-H-H-P-H-H-P-P-H-H-P-P-H-H)



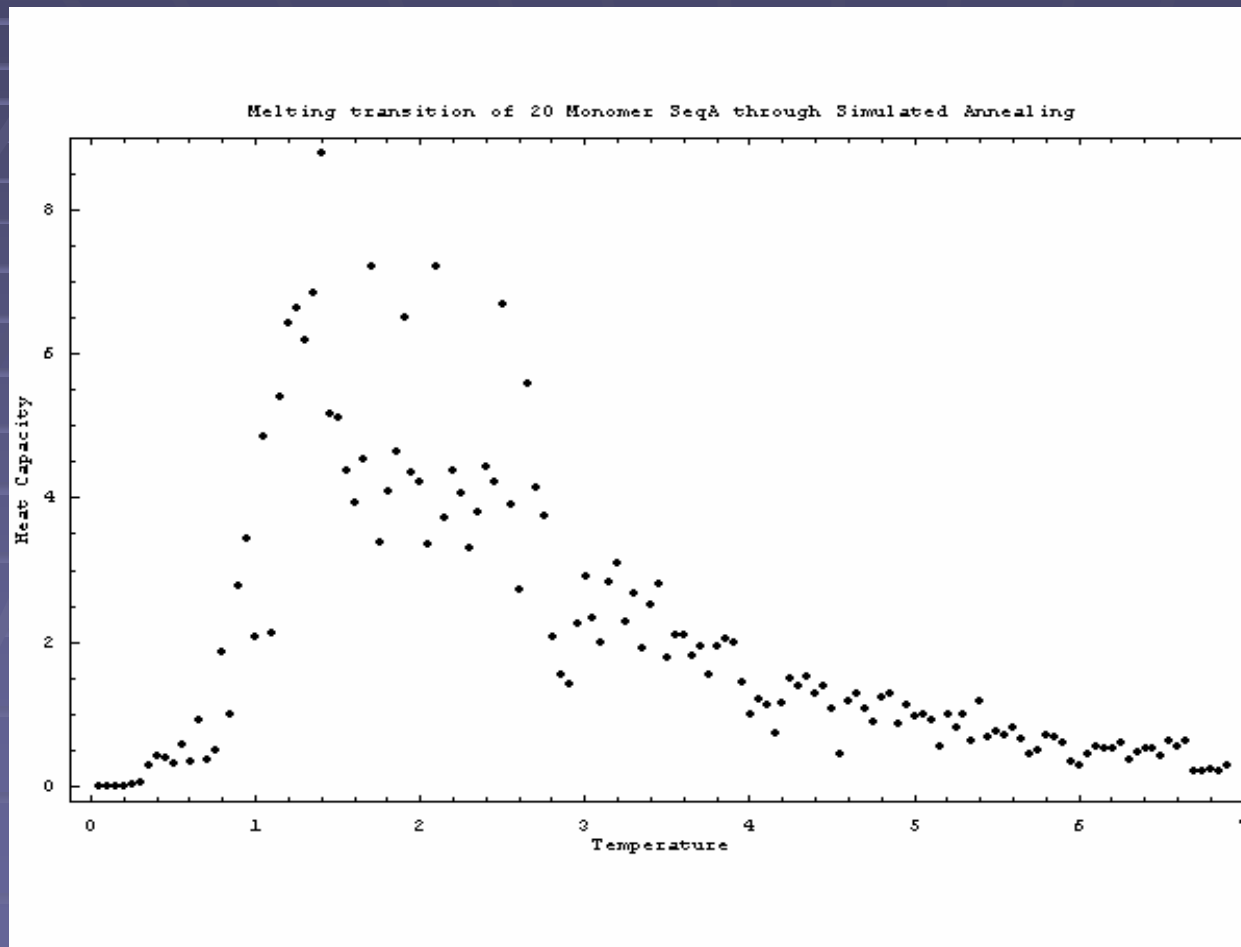
- Initial conformation ($E = -17.4$)



- Final conformation ($E = -26.4$)



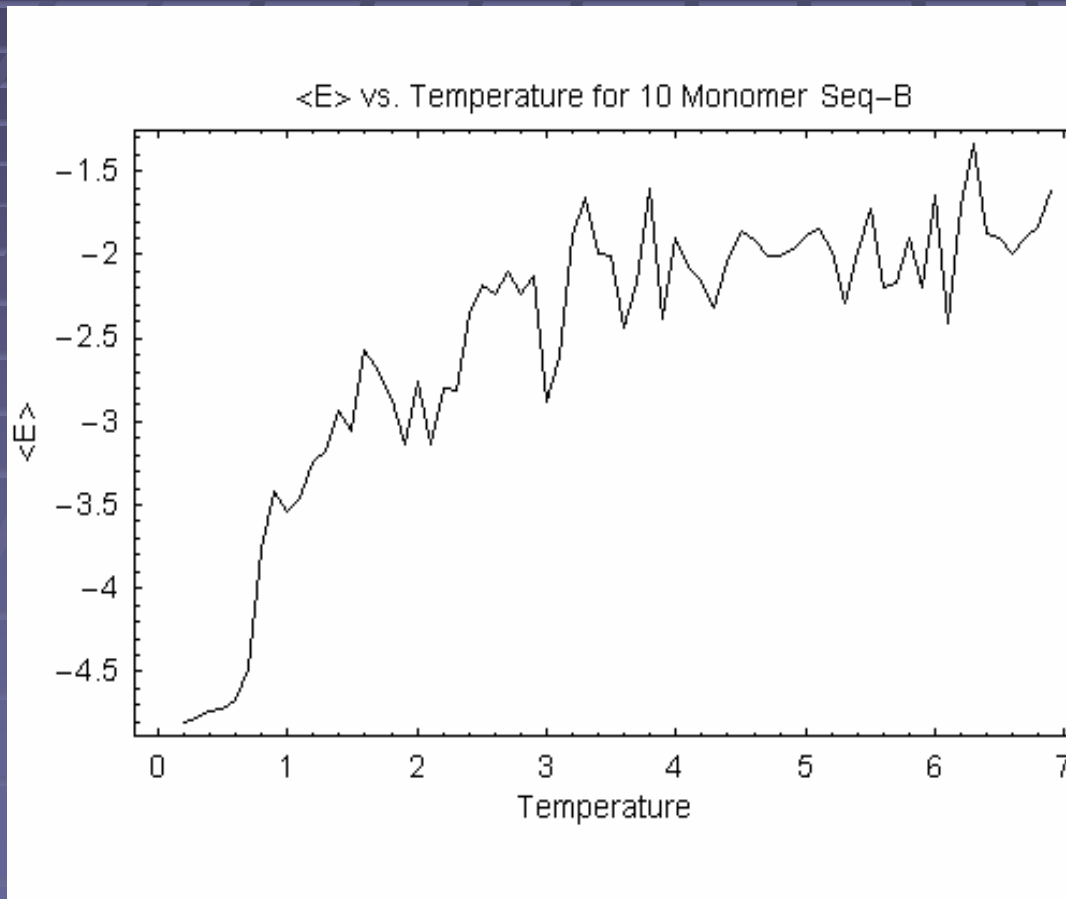
Results: 20 monomer Seq-A



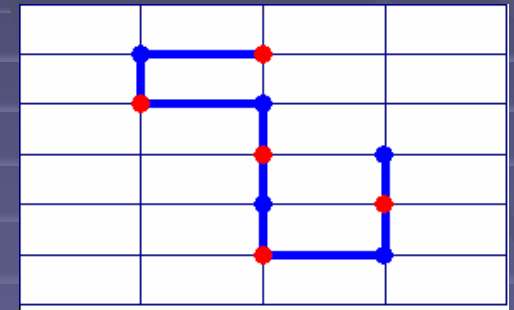
- Graph of $C(T)$ vs. T shows phase transition
- $T_c \approx 1.4$

Results: 10 Monomer Seq-B

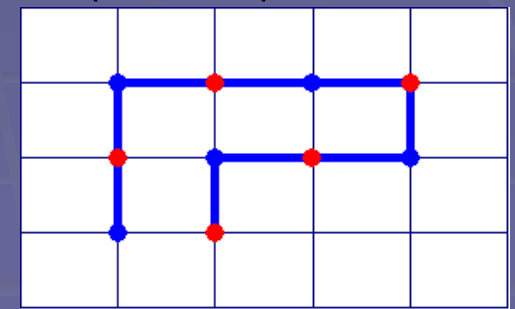
(H-P-H-P-H-P-H-P-H-P)



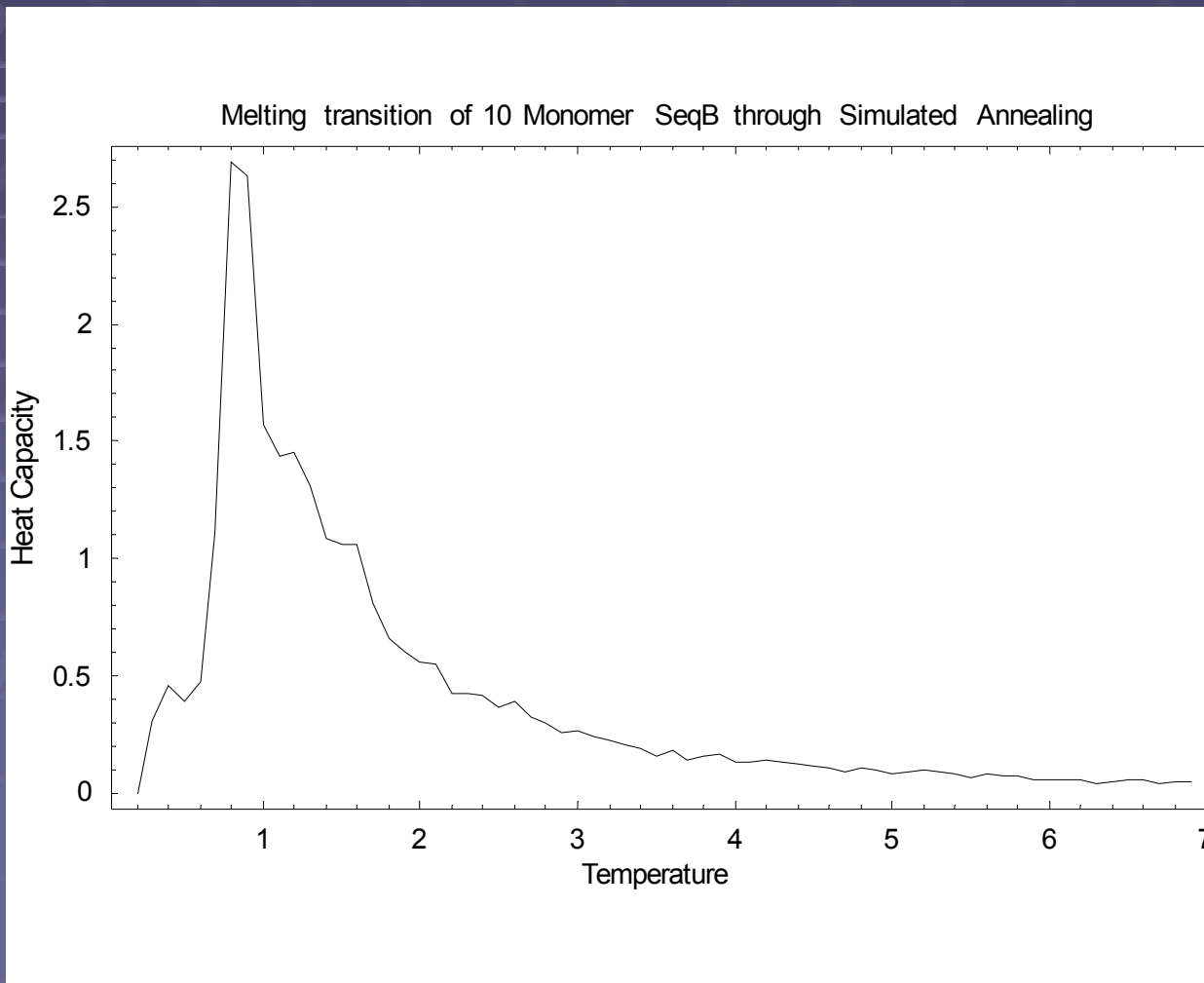
- Initial configuration ($E = -3.6$)



- Final configuration ($E = -4.8$)



Results: 10 Monomer Seq-B



■ $T_c \approx 1$

Conclusions

- Melting transitions were observed on plots of heat capacity vs. T
- Low energy conformations obtained
- Seq-A
 - observed to form possible 2D alpha helix conformation
 - Hydrophobic monomers arranged inwards
- Seq-B
 - Observed to form possible 2D beta-sheet

Future Work

- Increased Metropolis run-time (increased iterations)
- Comparison of low energy configurations for small sequences (<14 monomers) with lowest energy structure(s) from deterministic algorithm
- Implementation of 3-dimensional lattice model
- Comparison of 3-D folding structure with known native state structures
- Implementation of more effective reconfiguration moves

References

- [1] Metropolis, N.; Rosenbluth, A. W.; Rosenbluth, M. N.; Teller, A. H.; Teller, E. *Journal of Chemical Physics* (1953). 21: 1087-1092
- [2] Kirkpatrick, S; Gelatt, C. D.; Vecchi, M. P. *Science* (1983), Volume 220: 671-679.
- [3] Levinthal, C. University of Illinois Press (1968).
- [4] Leach, A. *Molecular Modeling: Principles and Applications*, 2nd Edition. Prentice Hall (2001): 423-431.
- [5] Dil, K. *Biochemistry* (1985), 24: 1501-1509.
- [6] Klimov, D. K.; Thirumalai, D. *Physical Review Letters* (1996), Volume 76: 4070-4073.
- [7] Bahar, I.; Atilgan, A. R.; Jernigan, R. L.; Erman, Burak. *Proteins: Structure, Function, and Genetics* (1997), 29: 172-185.
- [8] Rathore, Nitin; de Pablo, Juan. *Journal of Chemical Physics* (2002), Volume 116: 7225-7230.

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